







## CLINicell® Specifications

<b>Description :</b>	Fully closed and secured cell culture system intended to ensure the safety of both cell culture and manipulator in a controlled sterile environment. The rigid frame makes the handling easier and increase the manipulation safety. <b>CLINicell®</b> are stackable to provide economy in incubators.								
<b>Applications :</b>	<ul style="list-style-type: none"> <li>❖ Expansion of hematopoietic progenitors and stem cells,</li> <li>❖ Production of dendritic cells from monocytes and from CD34<sup>+</sup> cells,</li> <li>❖ Selection and expansion of mesenchymal stem cells,</li> <li>❖ Culture of hybridoma and production of antibodies,</li> <li>❖ Culture of immortal cell lines...</li> </ul>								
<b>Dimensions :</b>	<b>CLINicell® 25</b> 97 mm x 72 mm x 12 mm, <b>CLINicell® 250</b> 250 mm x 175 mm x 16 mm.								
<b>Culture area :</b>	<b>CLINicell® 25</b> 25 cm <sup>2</sup> x 2, <b>CLINicell® 250</b> 250 cm <sup>2</sup> x 2.								
<b>Volume :</b>	<b>CLINicell® 25</b> 5 ml to 10 ml, <b>CLINicell® 250</b> 80 ml to 160 ml.								
<b>Materials :</b>	Polycarbonate rigid frame, gas permeable polycarbonate films. Totally flat and transparent, the polycarbonate films are treated for cell culture and allow an excellent observation under optical microscopes.								
<b>Gas Transfer Rate* :</b>	<table border="0"> <tr> <td>Air</td> <td>85 ml/mil/100 in<sup>2</sup>/24 hr/atm</td> </tr> <tr> <td>Nitrogen</td> <td>50 ml/mil/100 in<sup>2</sup>/24 hr/atm</td> </tr> <tr> <td>Oxygen</td> <td>300 ml/mil/100 in<sup>2</sup>/24 hr/atm</td> </tr> <tr> <td>Carbon Dioxide</td> <td>1.075 ml/mil/100 in<sup>2</sup>/24 hr/atm</td> </tr> </table>	Air	85 ml/mil/100 in <sup>2</sup> /24 hr/atm	Nitrogen	50 ml/mil/100 in <sup>2</sup> /24 hr/atm	Oxygen	300 ml/mil/100 in <sup>2</sup> /24 hr/atm	Carbon Dioxide	1.075 ml/mil/100 in <sup>2</sup> /24 hr/atm
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<b>Connections :</b>	All types of systems with "Luer Lock" standard connections (syringes, filters...). The <b>CLINicell® 250</b> has an injection site. <b>CLINicell®</b> may be connected in parallel or series, together and to other systems such as bags and <b>CELLPerf®</b> , to allow different perfusion options.								
<b>User instructions :</b>	Available, in French or in English, in PDF format downloadable from website <a href="http://www.mabio.net">www.mabio.net</a>								

<b>Symbols :</b>	 Warning, see the joined documents
	 Irradiation sterilized
	 Do not reuse
	 Batch number
	 Expiry date
	 CE Mark by KEMA medical n° 2111774CE01

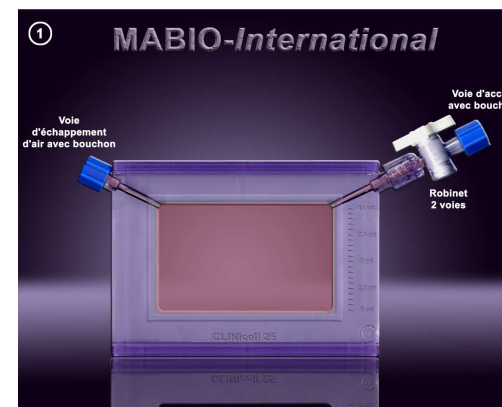
\* valeurs fournies par le fabricant

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Laboratoires  
**MABIO®**

## User Instructions

### CLINicell® 25 Sterile single use cell culture cassette.



**Please read this user manual prior to any manipulation**

**For additional information to assist manipulation of our products, please contact :**

**Tel : +33 (0)320 234 196 Fax. : +33 (0)320 234 086 E-mail : [mabio@mabio.net](mailto:mabio@mabio.net)**

#### I. Storage precautions:

Store in a clean, dry and dark location and at room temperature.

#### II. Handling precautions:

- a- Do not use **CLINicell®** if it is damaged. Do not reuse. Always verify packaging integrity. We guarantee **CLINicell®** sterility if packaging is unopened and intact.
- b- Manipulate **CLINicell®** in a sterile environment.  
Avoid contact between the access ports and your hands or the flow hood working surface. Check that all the ways are closed or connected before getting **CLINicell®** out of the flow hood.
- c- Before any manipulation, check that the stopcock connection (1) is secured and ensure the stopcock is closed.
- d- For any manipulation (injection or extraction of fluids) always open the ventilation port to avoid overpressure inside the **CLINicell®**.
- e- We suggest to use "Luer Lock" syringe, as this allows the syringe to securely screw to the access port.
- f- For safety, we recommend to use blunt-end needles to aspirate solutions with the syringe.
- g- To limit the contamination risks when you remove the syringe from the access port, always create a forced vacuum to draw any residual liquid remaining in the port end.

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## Manipulation of the **CLINicell® 25** Culture Cassette

We recommend to prepare the cell suspension within the optimal volume of medium (10 ml) and to inject this cell suspension into the **CLINicell®**.

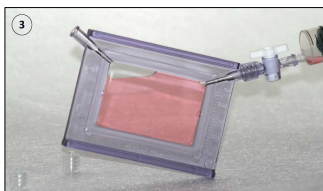
For high concentration cultures (from  $1.10^6$  cells/ml) of non-adherent cells, we recommend to re-suspend the cells each two days. Gently agitate or tap on the rigid frame of the **CLINicell®**.

### Filling the **CLINicell® 25**

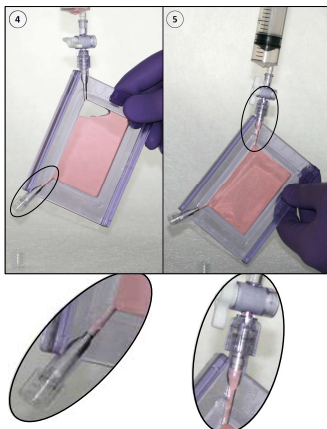
1.) Check that the stopcock connection (1) is secured and ensure it is closed.



- 2.a) Prepare the cell suspension in the optimal volume of medium (10 ml),
- 2.b) Stand the **CLINicell®** vertically,
- 2.c) Remove both caps,
- 2.d) Take cell suspension using a 20 ml syringe containing about 3 ml of air (2).



- 3.a) Connect the syringe to the access port and open the stopcock,
- 3.b) Inject the suspension (lift carefully the **CLINicell®** so that the ventilation port rises up),
- 3.c) Inject contained air in the syringe to push the suspension present in the stopcock (3).



- 4.) Tilt gently the **CLINicell®** so that the access point is raised up till the suspension slightly goes in the air exit port and screw its cap (4),
- 5.) Extract the residual air contained in the cassette and close the stopcock (5), then create a forced vacuum with the syringe and disconnect it from the access port.



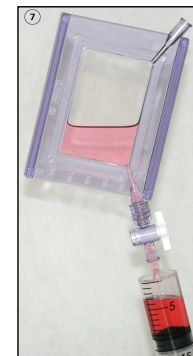
6.) Screw the access port cap and incubate the **CLINicell®** (6).

### Recovery of non-adherent cells

- 1.) Re-suspend the cells, gently agitate or gently tap on the rigid frame of the **CLINicell®**.
- 2.) Stand the **CLINicell®** vertically (1).
- 3.) Ensure the stopcock is closed and remove the access port cap.
- 4.) Connect a 20 ml syringe to the access port and open the stopcock.
- 5.) Remove the ventilation port cap.
- 6.) Take the **CLINicell®** in your hand to ensure that the ventilation port is raised up and aspirate the cell suspension (7)
- 7.) Secure the ventilation port cap and close the stopcock.
- 8.) Disconnect the syringe (create a forced vacuum) and secure the access port cap.

## Recommended washing procedure following cell recovery

- 1.) Stand the **CLINicell®** vertically (1),
- 2.) Ensure the stopcock is closed and remove the access port cap.
- 3.) Connect a 10 ml syringe containing the washing solution (e.g. 5 ml of PBS) and 2-3 ml of air and open the stopcock.
- 4.) Remove the ventilation port cap and inject the washing solution. During the injection, lift the **CLINicell®** so that the ventilation port is raised up to allow air to escape.
- 5.) Secure the ventilation port cap and gently agitate the **CLINicell®**.
- 6.) Stand the **CLINicell®** vertically and remove the ventilation port cap
- 7.) Take the **CLINicell®** in your hand to ensure the ventilation port is raised (pointing up), and aspirate the washing solution
- 8.) Replace the ventilation port cap and close the stopcock.
- 9.) Disconnect the syringe (create a forced vacuum) and secure the access port cap.



### Recovery of adherent cells

- 1.) Stand the **CLINicell®** vertically (1).
- 2.) Ensure the stopcock is closed and remove the access port cap.
- 3.) Connect a 20 ml syringe to the access port, open the stopcock and remove the ventilation port cap
- 4.) Take **CLINicell®** in your hand to ensure the ventilation port is raised up and aspirate the supernatant (7).
- 5.) Close the stopcock and disconnect the syringe (create a forced vacuum).
- 6.) Refer to the washing procedure as explained in the previous section.
- 7.) Stand the **CLINicell®** vertically (1).
- 8.) Connect to the access port a 10 ml syringe containing the dissociation solution (e.g. 3 ml of trypsin /EDTA) and 2-3 ml of air.
- 9.) Open the stopcock, remove and inject the dissociation solution.
- 10.) Replace the ventilation port cap and close the stopcock.
- 11.) Disconnect the syringe (create a forced vacuum) and secure the access port cap.
- 12.) Gently agitate the **CLINicell®** to homogenize the repartition of the solution.
- 13.) If necessary, incubate at 37°C.
- 14.) Stand the **CLINicell®** vertically (1).
- 15.) Connect to the access port a 10 ml syringe containing the medium (or another solution) used to stop the dissociation reaction and 2-3 ml of air.
- 16.) Open the stopcock and remove the ventilation port cap.
- 17.) Lift the syringe and inject the medium.
- 18.) Replace the ventilation port cap and gently agitate the **CLINicell®** to homogenize.
- 19.) Stand the **CLINicell®** vertically (1) and remove the ventilation port cap.
- 20.) Take **CLINicell®** in your hand to ensure the ventilation port is raised up and aspirate the suspension (7).
- 21.) Secure the ventilation port cap and close the stopcock.
- 22.) Disconnect the syringe (create a forced vacuum) and secure the access port cap.
- 23.) If necessary, refer to the washing procedure as explained in the previous section
- 24.) Throw the used **CLINicell®** in an appropriate bin.